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MICROWAVE METHOD FOR PREPARING RADIOLABELLED GALLIUM COMPLEXES

The present invention relates to a method of producing radiolabelled gallium complexes. The complexes could be used as diagnostic agents, e.g. for positron emission tomography (PET) imaging.

PET imaging is a tomographic nuclear imaging technique that uses radioactive tracer molecules that emit positrons. When a positron meets an electron, the both are annihilated and the result is a release of energy in form of gamma rays, which are detected by the PET scanner. By employing natural substances that are used by the body as tracer molecules, PET does not only provide information about structures in the body but also information about the physiological function of the body or certain areas therein. A common tracer molecule is for instance 2-fluoro-2-deoxy-D-glucose (FDG), which is similar to naturally occurring glucose, with the addition of a ¹⁸Fatom. Gamma radiation produced from said positron-emitting fluorine is detected by the PET scanner and shows the metabolism of FDG in certain areas or tissues of the body, e.g. in the brain or the heart. The choice of tracer molecule depends on what is being scanned. Generally, a tracer is chosen that will accumulate in the area of interest, or be selectively taken up by a certain type of tissue, e.g. cancer cells. Scanning consists of either a dynamic series or a static image obtained after an interval during which the radioactive tracer molecule enters the biochemical process of interest. The scanner detects the spatial and temporal distribution of the tracer molecule. PET also is a quantitative imaging method allowing the measurement of regional concentrations of the radioactive tracer molecule.

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Commonly used radionuclides in PET tracers are ¹¹C, ¹⁸F, ¹⁵O ¹³N or ⁷⁶Br. Recently, new PET tracers were produced that are based on radiolabelled metal complexes comprising a bifunctional chelating agent and a radiometal. Bifunctional chelating agents are chelating agents that coordinate to a metal ion and are linked to a targeting vector that will bind to a target site in the patient's body. Such a targeting vector may be a peptide that binds to a certain receptor, probably associated with a certain area in the body or with a certain disease. A targeting vector may also be an oligonucleotide specific for e.g. an activated oncogene and thus aimed for tumour localisation. The advantage of such complexes is that the bifunctional chelating PZ0334/FV11.04.2003

agents may be labelled with a variety of radiometals like, for instance, ⁶⁸Ga, ²¹³Bi or ⁸⁶Y. In this way, radiolabelled complexes with special properties may be "tailored" for certain applications.

- 5 ⁶⁸Ga is of special interest for the production of Ga-radiolabelled metal complexes used as tracer molecules in PET imaging. ⁶⁸Ga is obtained from a ⁶⁸Ge/⁶⁸Ga generator, which means that no cyclotron is required. ⁶⁸Ga decays to 89% by positron emission of 2.92 MeV and its 68 min half life is sufficient to follow many biochemical processes *in vivo* without unnecessary radiation. With its oxidation state of +III, ⁶⁸Ga forms stable complexes with various types of chelating agents and ⁶⁸Ga tracers have been used for brain, renal, bone, blood pool, lung and tumour imaging.
 - J. Schumacher et al., Cancer Res. 61, 2001, 3712-3717 describe the synthesis of ⁶⁸Ga-N,N [2-hydroxy-5-(ethylene-β-carboxy)benzyl]ethylenediamine-N,N'-diacetic acid (⁶⁸Ga-HBED-CC). ⁶⁸Ga obtained from a ⁶⁸Ge/⁶⁸Ga generator and Ga³⁺ carrier are reacted with the chelating agent HBED-CC in acetate buffer for 15 min at 95°C. Uncomplexed ⁶⁸Ga is separated from the complex using a cation exchange column. The overall preparation is reported to take 70 min. A disadvantage of this method is that the overall preparation time of the radiolabelled complex is very long. Due to the addition of "cold" Ga³⁺ carrier, the specific activity of the reaction is low. Moreover, the radiolabelled complex had to be purified after the complex formation reaction.

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- WO-A-99/56791 discloses the reaction of ⁶⁸GaCl₃ obtained from a ⁶⁸Ge/⁶⁸Ga generator with the tetradentate amine trithiolate chelating agent tris(2-mercaptobenzyl)amine (S₃N). The complex formation is carried out at room temperature for 10 min. A disadvantage of the method described is that the radiolabelled complex had to be purified by liquid chromatography before it could be used for *in vivo* studies. A further disadvantage of the method is the relatively long reaction time
- Ö. Ugur et al., Nucl. Med. Biol. 29, 2002, 147-157 describe the synthesis of the ⁶⁸Ga labelled somatostatin analogue DOTA-DPhe¹-Tyr³-octreotide (DOTATOC). The compound is prepared by reacting ⁶⁸GaCl₃ obtained from a ⁶⁸Ge/⁶⁸Ga generator with

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the chelating agent DOTATOC for 15 min at 100°C. A disadvantage of this method is that the reaction mixture had to be heated at relatively high temperatures. The DOTA chelating agent was functionalised with a peptide targeting vector and peptides and proteins are substances, which are known to be sensitive to heat. Thus, with the method described there is a risk that heat sensitive targeting vectors are destroyed during complex formation. A further disadvantage is that the complex had to be purified by HPLC before it could be used for animal studies.

US-A-5070346 discloses ⁶⁸Ga-labelled complexes of the chelating agent tetraethylcyclohexyl-bis-aminoethanethiol (BAT-TECH). The complexes are synthesised by reacting ⁶⁸GaCl₃ obtained from a ⁶⁸Ge/⁶⁸Ga generator with BAT-TECH at 75°C for 15 min and subsequent filtration. The preparation of the complex was accomplished in 40 min. Due to the high reaction temperature; this method would not be suitable for bifunctional chelating agents comprising a heat sensitive targeting vector, for instance a peptide or a protein. A further disadvantage is the long reaction time of the complex formation reaction.

In view of the relatively short half-life of ⁶⁸Ga there is a need for a fast method for the synthesis of ⁶⁸Ga-labelled complexes, which could be used as tracer molecules for PET imaging.

It has now been found that the use of microwave activation substantially improves the efficiency and reproducibility of the ⁶⁸Ga-chelating agent complex formation. Due to microwave activation, chemical reaction times could be shortened substantially; i.e. the reaction is completed within 2 min and less. This is a clear improvement as a 10 minutes shortage of the reaction time saves about 10% of the ⁶⁸Ga activity. Furthermore, microwave activation also leads to fewer side reactions and to an increased radiochemical yield, which is due to increased selectivity. Solutions of ⁶⁶Ga³⁺, ⁶⁷Ga³⁺ and ⁶⁸Ga³⁺ radioisotopes, which have been obtained by cyclotron production or from a generator contain so-called pseudo carriers, i.e. other metal cations like for instance Fe³⁺, Al³⁺, Cu²⁺, Zn²⁺ and In³⁺. As these pseudo carriers compete with Ga³⁺ in the complex formation reaction, it is important to increase the selectivity of the radiolabelling reaction. Hence, microwave activation

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has a positive effect on radiolabelling with all Ga-radioisotopes, namely with ⁶⁶Ga, ⁶⁷Ga and ⁶⁸Ga.

- Microwave activation has been used in nucleophilic aromatic radiofluorations with ¹⁸F and it was found that comparable or better yields than those reported for thermal treatments were obtained in shorter reaction times (S. Stone-Elander et al., Appl. Rad. Isotopes 44(5), 1993, 889-893). However, the use of microwave activation in Ga-radiolabelling reactions has not been described yet.
- 10 The invention thus provides a method of producing a radiolabelled gallium complex by reacting a Ga³⁺ radioisotope with a chelating agent characterised in that the reaction is carried out using microwave activation.
- Suitable Ga³⁺ radioisotopes according to the invention are ⁶⁶Ga³⁺, ⁶⁷Ga³⁺ and ⁶⁸Ga³⁺, preferably ⁶⁶Ga³⁺ and ⁶⁸Ga³⁺ and particularly preferably ⁶⁸Ga³⁺. ⁶⁶Ga³⁺ and ⁶⁸Ga³⁺ are particularly suitable for the production of radiolabelled complexes useful in PET imaging whereas ⁶⁷Ga³⁺ is particularly suitable for the production of radiolabelled complexes useful in single photon emission computerised tomography (SPECT).
- 20 ⁶⁶Ga³⁺ is obtainable by cyclotron production by irradiation of elemental zinc targets. To minimise the amounts of ⁶⁷Ga production, the target thickness is preferably maintained such that the degraded proton energy is above 8 MeV, and irradiation time is kept short, e.g. <4 hrs. The chemical separation may be achieved using solvent-solvent extraction techniques using isopropyl ether and HCl as described in L.C. Brown, Int. J. Appl. Radiat. Isot. 22, 1971, 710-713. ⁶⁶Ga has a relatively long half-life of 9.5 h and the most abundant positron emitted has a uniquely high energy of 4.2 MeV.
- ⁶⁷Ga³⁺ is obtainable by cyclotron production and ⁶⁷GaCl₃ obtained by cyclotron production is a commercially available compound. The half-life of ⁶⁷Ga is 78 h.
 - ⁶⁸Ga is obtainable from a ⁶⁸Ge/⁶⁸Ga generator. Such generators are known in the art and for instance described by C. Loc'h et al, J. Nucl. Med. 21, 1980, 171-173. Generally, ⁶⁸Ge is loaded onto a column consisting of an organic resin or an

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inorganic metal oxide like tin dioxide, aluminium dioxide or titanium dioxide. ⁶⁸Ga is eluted from the column with aqueous HCl, yielding ⁶⁸GaCl₃. ⁶⁸Ga³⁺ is particularly preferred in the method according to the invention as its production does not re-quire a cyclotron and its 68 min half-life is sufficient to follow many biochemical processes *in vivo* by PET imaging without long radiation.

Preferred chelating agents for use in the method of the invention are those which present the Ga³⁺ radioisotopes in a physiologically tolerable form. Further preferred chelating agents are those that form complexes with Ga³⁺ radioisotopes that are stable for the time needed for diagnostic investigations using the radiolabelled complexes.

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Suitable chelating agents are, for instance, polyaminopolyacid chelating agents like DTPA, EDTA, DTPA-BMA, DOA3, DOTA, HP-DOA3, TMT or DPDP. Those chelating agents are well known for radiopharmaceuticals and radiodiagnosticals. Their use and synthesis are described in, for example, US-A-4647447, US-A-5362 475, US-A-5534241, US-A-5358704, US-A-5198208, US-A-4963344, EP-A-230893, EP-A-130934, EP-A-606683, EP-A-438206, EP-A-434345, WO-A-97/00087, WO-A-96/40274, WO-A-96/30377, WO-A-96/28420, WO-A- 96/16678, WO-A-96/11023, WO-A-95/32741, WO-A-95/27705, WO-A-95/26754, WO-A-95/28967, WO-A-95/28392, WO-A-95/24225, WO-A-95/17920, WO-A-95/15319, WO-A-95/09848, WO-A-94/27644, WO-A-94/22368, WO-A-94/08624, WO-A-93/16375, WO-A-93/06868, WO-A-92/11232, WO-A-92/09884, WO-A-92/08707, WO-A-91/15467, WO-A-91/10669, WO-A-91/10645, WO-A-91/07191, WO-A-91/05762, WO-A-90/12050, WO-A-90/03804, WO-A-89/00052, WO-A-89/00557, WO-A-88/01178, WO-A-86/02841 and WO-A-86/02005.

Suitable chelating agents include macrocyclic chelating agents e.g. porphyrin-like molecules and pentaaza-macrocycles as described by Zhang et al., Inorg. Chem. 37(5), 1998, 956-963, phthalocyanines, crown ethers, e.g. nitrogen crown ethers such as the sepulchrates, cryptates etc., hemin (protoporphyrin IX chloride), heme and chelating agents having a square-planar symmetry.

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Macrocyclic chelating agents are preferably used in the method of the invention. In a preferred embodiment, these macrocyclic chelating agents comprise at least one hard donor atom such as oxygen and/or nitrogen like in polyaza- and polyoxomacrocycles. Preferred examples of polyazamacrocyclic chelating agents include DOTA, TRITA, TETA and HETA with DOTA being particularly preferred.

Particularly preferred macrocyclic chelating agents comprise functional groups such as carboxyl groups or amine groups which are not essential for coordinating to Ga³⁺ and thus may be used to couple other molecules, e.g. targeting vectors, to the chelating agent. Examples of such macrocyclic chelating agents comprising functional groups are DOTA, TRITA or HETA.

In a further preferred embodiment, bifunctional chelating agents are used in the method according to the invention. "Bifunctional chelating agent" in the context of the invention means chelating agents that are linked to a targeting vector. Suitable targeting vectors for bifunctional chelating agents useful in the method according to the invention are chemical or biological moieties, which bind to target sites in a patient's body, when the radiolabelled gallium complexes comprising said targeting vectors have been administered to the patient's body. Suitable targeting vectors for bifunctional chelating agents useful in the method according to the invention are proteins, glycoproteins, lipoproteins, polypeptides like antibodies or antibody fragments, glycopolypeptides, lipopolypeptides, peptides, like RGD binding peptides, glycopeptides, lipopoptides, carbohydrates, nucleic acids e.g. DNA, RNA, oligonucleotides like antisense oligonucleotides or a part, a fragment, a derivative or a complex of the aforesaid compounds, or any other chemical compound of interest like relatively small organic molecules, particularly small organic molecules of less than 2000 Da.

In a particularly preferred embodiment, macrocyclic bifunctional chelating agents are used in the method according to the invention. Preferred macrocyclic bifunctional chelating agents comprise DOTA, TRITA or HETA linked to a targeting vector, preferably to a targeting vector selected from the group consisting of proteins, glycoproteins, lipoproteins, polypeptides, glycopolypeptides, lipopolypeptides, peptides, glycopeptides, lipopeptides carbohydrates, nucleic acids,

oligonucleotides or a part, a fragment, a derivative or a complex of the afor esaid compounds and small organic molecules; particularly preferably to a targeting vector selected from the group consisting of peptides and oligonucleotides.

5 The targeting vector can be linked to the chelating agent via a linker group or via a spacer molecule. Examples of linker groups are disulfides, ester or amides, examples of spacer molecules are chain-like molecules, e.g. lysin or hexylamine or short peptide-based spacers. In a preferred embodiment, the linkage between the targeting vector and the chelating agent part of radiolabelled gallium complex is as such that the targeting vector can interact with its target in the body without being blocked or hindered by the presence of the radiolabelled gallium complex.

Microwave activation according to the invention is suitably carried out by using a microwave oven, preferably by using a monomodal microwave oven as. Suitably microwave activation is carried out at 80 to 120 W, preferably at 90 to 110 W, particularly preferably at about 100 W. Suitable microwave activation times range from 20 s to 2 min, preferably from 30 s to 90 s, particularly preferably from 45 s to 60 s.

- A temperature control of the reaction is advisable when temperature sensitive chelating agents, like for instance bifunctional chelating agents comprising peptides or proteins as targeting vectors, are employed in the method according to the invention. Duration of the microwave activation should be adjusted in such a way, that the temperature of the reaction mixture does not lead to the decomposition of the chelating agent and/or the targeting vector. If chelating agents used in the method according to the invention comprise peptides or proteins, higher temperatures applied for a shorter time are generally more favourable than lower temperatures applied for a longer time period.
- Microwave activation can be carried out continuously or in several microwave activation cycles during the course of the reaction.

In a preferred embodiment, the invention provides a method of producing a ⁶⁸Ga radiolabelled PET imaging tracer by reacting ⁶⁸Ga³⁺ with a macrocyclic bifunctional

chelating agent comprising hard donor atoms, characterised in that the reaction is carried out using microwave activation.

In a particularly preferred embodiment of the method described in the last preceding paragraph, the microwave activation is carried out from 30 s to 90 s at 90 to 110 W.

If ⁶⁸Ga³⁺ is used in the method according to the invention, the ⁶⁸Ga³⁺ is preferably obtained by contacting the eluate form a ⁶⁸Ge/⁶⁸Ga generator with an amion exchanger and eluting ⁶⁸Ga³⁺ from said anion exchanger. In a preferred embodimment, the anion exchanger is an anion exchanger comprising HCO₃ as counterions.

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The use of anion exchangers to treat ⁶⁸Ga eluate obtained from a ⁶⁸Ge/⁶⁸Ga generator is described by J. Schuhmacher et al. Int. J. appl. Radiat. Isotopes 32, 1981, 31-36. A Bio-Rad AG 1 x 8 anion exchanger was used for treating the 4.5 N HCl ⁶⁸Ga eluate obtained from a ⁶⁸Ge/⁶⁸Ga generator in order to decrease the amount of ⁶⁸Ge present in the eluate.

It has now been found that the use of anion exchangers comprising HCO₃ as counterions is particularly suitable for the purification and concentration of the generator eluate. Not only the amount of ⁶⁸Ge present in the eluate could be reduced but also the amount of so-called pseudo carriers, i.e. other metal cations like Fe³⁺, Al3+, Cu2+, Zn2+ and In3+, that are eluted together with the 68Ga3+ from the generator. As these pseudo carriers compete with ⁶⁸Ga³⁺ in the subsequent complex formation reaction, it is especially favourable to reduce the amount of those cations as much as possible before the labelling reaction. A further advantage of the anion-exchange purification step is that the concentration of ⁶⁸Ga³⁺, which is in the picomolar to nanomolar range after the elution, can be increased up to a nanomolar to micromolar level. Hence, it is possible to reduce the amount of chelating agent in a subsequent complex formation reaction, which considerably increases the specific radioactivity. This result is important for the production of ⁶⁸Ga-radiolabelled PET tracers that comprise a bifunctional chelating agent; i.e. a chelating agent linked to a targeting vector, as the increase in specific radioactivity enables the reduction in amount of such tracers when used in a patient.

Hence, another preferred embodiment of the method according to the invention is a method of producing a ⁶⁸Ga- radiolabelled complex by reacting ⁶⁸Ga³⁺ with a chelating agent using microwave activation, wherein the ⁶⁸Ga³⁺ is obtained by contacting the cluate form a ⁶⁸Ge/⁶⁸Ga generator with an anion exchanger, preferably with an anion exchanger comprising HCO₃ as counterions, and cluting ⁶⁸Ga³⁺ from said anion exchanger.

⁶⁸Ge/⁶⁸Ga generators are known in the art, see for instance C. Loc'h et al, J. Nucl. Med. 21, 1980, 171-173 or J. Schuhmacher et al. Int. J. appl. Radiat. Isotopes 32, 1981, 31-36. ⁶⁸Ge may be obtained by cyclotron production by irradiation of, for instance Ga₂(SO₄)₃ with 20 MeV protons. It is also commercially available, e.g. as ⁶⁸Ge in 0.5 M HCl. Generally, ⁶⁸Ge is loaded onto a column consisting of organic resin or an inorganic metal oxide like tin dioxide, aluminium dioxide or titarnium dioxide. ⁶⁸Ga is eluted from the column with aqueous HCl yielding ⁶⁸GaCl₃.

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Suitable columns for ⁶⁸Ge/⁶⁸Ga generators consist of inorganic oxides like aluminium dioxide, titanium dioxide or tin dioxide or organic resins like resins comprising phenolic hydroxyl groups (US-A-4264468) or pyrogallol (J. Schuhmacher et al., Int. J. appl. Radiat. Isotopes 32, 1981, 31-36). In a preferred embodiment, a ⁶⁸Ge/⁶⁸Ga generator comprising a column comprising titanium dioxide is used in the method according to the invention.

The concentration of the aqueous HCl used to elute the ⁶⁸Ga from the ⁶⁸Ge/⁶⁸Ga generator column depends on the column material. Suitably 0.05 to 5 M HCl is used for elution of ⁶⁸Ga. In a preferred embodiment, the eluate is obtained from a ⁶⁸Ge/⁶⁸Ga generator comprising a column comprising titanium dioxide and ⁶⁸Ga is eluted using 0.05 to 0.1 M HCl, preferably about 0.1 M HCl.

In a preferred embodiment of the method according to the invention, a strong anion exchanger comprising HCO₃ as counterions, preferably a strong anion exchanger comprising HCO₃ as counterions, is used. In a further preferred embodiment, this anion exchanger comprises quaternary amine functional groups. In another further preferred embodiment, this anion exchanger is a strong anion exchange resin based on polystyrene-divinylbenzene. In a particularly preferred embodiment, the anion

exchanger used in the method according to the invention is a strong anion exch ange resin comprising HCO₃ as counterions, quaternary amine functional groups and the resin is based on polystyrene-divinylbenzene.

Suitably, water is used to elute the ⁶⁸Ga from the anion exchanger in the me-thod according to the invention.

Examples

5 Example 1:

Comparison of ⁶⁸Ga – radiolabelling of DOTA-D-Phe¹-Tyr³ – Octrectide (DOTA-TOC) using conventional heating and microwave activation:

1a) ⁶⁸Ga – radiolabelling of DOTA-TOC using conventional heating:

Sodium acetate was added to the eluate from a ⁶⁸Ge/⁶⁸Ga-generator (36 mg to 1 mL) to adjust the pH of the eluate to approximately 5.5 and the mixture was vortexed well. DOTA-TOC (20 nmol) was added and the reaction mixture was heated at 96 °C for 25 min. The reaction mixture was cooled to room temperature and applied to a C-18 SPE-column (HyperSEP S C18), which was then washed with 2 mL H₂O and the product was eluted with ethanol: water 50:50 (1 mL).

The reaction mixture and the product were analysed by HPLC using Vydac RP and Fast Desalting HR 10/10 FPLC gel filtration columns.

The analytical radiochemical yield (RCY) was 67%.

20 The isolated RCY was 34%.

Electrospray ionization mass spectrometry, ESI-MS, was performed on Fisons Platform (Micromass, Manchester, UK), using positive mode scanning and detecting $[M+2H]^{2+}$. DOTATOC was detected at m/z = 711.26 and authentic Ga-DOTATOC was detected at m/z = 746.0 (calculated m/z = 746.5).

1b) ⁶⁸Ga – radiolabelling of DOTA-TOC using microwave activation

The reaction mixture was prepared identically as described under 1a) and transferred into a Pyrex glass vial for microwave activation for 1 min at 100 W. The reaction mixture was cooled to room temperature and applied to a C-18 SPE-column (HyperSEP S C18), which was then washed with 2 mL H₂O and the product was eluted with ethanol: water 50:50 (1 mL).

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The reaction mixture and the product were analysed by HPLC using Vydac RP and Fast Desalting HR 10/10 FPLC gel filtration columns.

5 The analytical RCY was over 98%.

The isolated RCY was 70%.

Electrospray ionization mass spectrometry, ESI-MS, was performed on Fi sons Platform (Micromass, Manchester, UK), using positive mode scanning and detecting $[M+2H]^{2+}$. DOTATOC was detected at m/z = 711.26 and authentic Ga-DOTATOC was detected at m/z = 746.0 (calculated m/z = 746.5).

15 1c) Results of the comparison

In the case of microwave activation, the amount of radioactive material and the product specific activity was increased by 21%. The isolated radiochemical yield was increased 2 fold compared to the results obtained with conventional heating. As the radiochemical yield of the reaction mixture in case of microwave activation was over 98%, a further purification would not have been necessary and the crude reaction mixture could have been used for *in vivo* application.

Example 2:

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25 ⁶⁸Ga radiolabelling of DOTA linked to oligonucleotides

In a first step, four different antisense oligonucleotides specific for activated human K-ras oncogene were linked to DOTA:

- 17-mer phosphodiester oligonucleotide with hexylaminolinker at 5' end;
- 17-mer phosphodiester oligonucleotide with hexylaminolinker at 3' end;
 - 17-mer phosphorothioate oligonucleotide with hexylaminolinker at 5' end; and
 - 2'-O-methyl phosphodiester with hexylaminolinker at 5' end.

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2a) Conjugation of DOTA to oligonucleotides:

DOTA (32 mg, 66 µmol) and Sulfo-NHS (14 mg, 65 µmol) in H₂O (250 µl) were added to EDC (13 mg, 68 µmol) in H₂O (250 µl), stirred on ice for 30 min and then warmed to room temperature to give DOTA-sulfo-NHS. A 100 fold excess of DOTA-NHS solution was added dropwise to the oligonucleotide (70-450 nmol) in 1M carbonate buffer (pH 9) and then cooled on ice. The mixture was left at room temperature for 10 hours. The reaction mixture was first purified by gel filtration with NAP 5 columns, eluted with H₂O and 100 μL of 1M TEAA (triethylammonium acetate buffer) was added to 1 mL of the product eluate. The product eluate was then applied to a C-18 SPE column (Supelco), the column was washed with 50 mM TEAA (5 mL), 50 mM TEAA containing 5% acetonitrile (3 mL) and the DOTAoligonucleotide was eluted with water:acetonitrile 50:50 (1 mL). The wateracetonitrile fraction was dried using a vacuum centrifuge. The products were analysed using electrospray ionization mass spectrometry. Analysis in negative mode after direct infusion resulted in the following data: 1. DOTA-phosphodiester: MS (ESI) m/z: 662.27 [M-8H]⁸; 756.36 [M-7H]⁷; 882.91 [M-6H]⁶. Reconstitution of the data gave M = 5303.71; 2. DOTA-phosphorotioate: MS (ESI) m/z: 656.58 [M- $8H_1^{9}$; 738.56 [M-7H]⁸. Reconstitution of the data gave M = 5917.35; 3. DOTA-2'-O-methyl phosphodiester: MS (ESI) m/z: 674.02 [M-6H]⁹; 770.19 [M-8H]⁸; 885.00 $[M-7H]^7$. Reconstitution of the data gave M = 6148.84

2b) ⁶⁸Ga – radiolabelling

Sodium acetate was added to the eluate from a ⁶⁸Ge/⁶⁸Ga-generator (36 mg to 1 mL) to adjust the pH of the eluate to approximately 5.5 and the mixture was vortexed well. DOTA-oligonucleotide (10-100 nmol) was added and the mixture was transferred into a Pyrex glass vial for microwave activation for 1 min at 100 W. The reaction mixture was cooled to room temperature then 1 mL of 150 mM TEAA in H₂O was added. The mixture was applied to a C-18 SPE-column (Supelco), which was then washed with 50 mM TEAA (1 mL), 50mM TEAA containing 5% acetonitrile (1mL). The product was eluted with ethanol: water 50:50 (1 mL) or water:acetonitrile 50:50 (1 mL). The reaction mixture was analysed by HPLC using Vydac RP and Fast Desalting HR 10/10 FPLC gel filtration columns. The analytical RCY ranged from 50% to 70%, the isolated RCY ranged from 30 to 52%. Larger amounts of stronger eluents might improve the isolated RCY.

Example 3:

⁶⁸Ga radiolabelling of DOTA linked to peptides

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In a first step, four different peptides were linked to DOTA:

- Vasoactive Intestinal Peptide (VIP); 28 amino acid residues:
- Neuropeptide Y Fragment 18-36 (NPY); 19 amino acid residues;
- Pancreastatin Fragment 37-52 (P); 16 amino acid residues; and
- Angiotensin II (A); 8 amino acid residues.

3a) Conjugation of DOTA to peptides:

Conjugation was carried out as described in 2a) using peptides (0.5-3 μ mol) instead of oligonucleotides.

The reaction mixtures and products were analysed by HPLC using Vydac RP and Fast Desalting HR 10/10 FPLC gel filtration columns. Electrospray ionization mass spectrometry, ESI-MS, was performed on Fisons Platform (Micromass, Manchester, UK), using positive mode scanning and detecting [M+2H]²⁺, [M+4H]⁴⁺ and [M+5H]⁵⁺. VIP was detected at m/z = 832.07 [M+4H]⁴⁺. (DOTA)₂-VIP was detected at m/z = 1025.00 [M+4H]⁴⁺. (DOTA)₃-VIP was detected at m/z = 1122.0 [M+4H]⁴⁺. (DOTA)₄-VIP was detected at m/z = 1218.00 [M+4H]⁴⁺. NPY was detected at m/z = 819.31 [M+3H]³⁺. DOTA-NPY was detected at m/z = 948.18 [M+3H]³⁺. P was detected at m/z = 909.55 [M+2H]²⁺. DOTA-P was detected at m/z = 1103.02 [M+2H]²⁺. A was detected at m/z = 524.1 [M+2H]²⁺ and DOTA-A was detected at m/z = 717.20 [M+2H]²⁺.

3b) ⁶⁸Ga – radiolabelling

⁶⁸Ga – radiolabelling was carried out as described in 2b) using 10-20 nmol DOTA-peptide.

The reaction mixture was analysed by HPLC using Vydac RP and Fast Desalting HR 10/10 FPLC gel filtration columns. The analytical RCY ranged from 80% to 90%,

the isolated RCY ranged from 60 to 70%. Larger amounts of stronger eluents ranight improve the isolated RCY.

Claims:

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- 1. Method of producing a radiolabelled gallium complex by reacting a Ga³⁺ radioisotope with a chelating agent characterised in that the reaction is carried out using microwave activation.
- 2. Method according to claim 1 wherein the Ga³⁺ radioisotope is selected from the group consisting of ⁶⁶Ga³⁺, ⁶⁷Ga³⁺ and ⁶⁸Ga³⁺.
- 3. Method according to claims 1 and 2 wherein the Ga³⁺ radioisotope is ⁶⁸Ga³⁺.
 - 4. Method according to claims 1 to 3 wherein the chelating agent is a macrocyclic chelating agent.
- Method according to claims 1 to 4 wherein the chelating agent comprises hard donor atoms, preferably O and N atoms.
 - 6. Method according to claims 1 to 5 wherein the chelating agent is a bifunctional chelating agent.

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- 7. Method according to claims 1 to 6 wherein the chelating agent is a bifunctional chelating agent comprising a targeting vector selected from the group consisting of proteins, glycoproteins, lipoproteins, polypeptides, glycopolypeptides, lipopolypeptides, peptides, glycopeptides, lipopeptides, carbohydrates, nucleic acids, oligonucleotides or a part, a fragment, a derivative or a complex of the aforesaid compounds and small organic molecules.
- 8. Method according to claim 7 wherein the target vector is a peptide or oligonucleotide.

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9. Method according to claims 1 to 8 wherein the microwave activation is carried out at 80 to 120 W, preferably at 90 to 110 W.

- 10. Method according to claims 1 to 9 wherein the microwave activation is carried out for 20 s to 2 min, preferably for 30 s to 90 s.
- 11. Method according to claims 3 to 10 wherein the ⁶⁸Ga³⁺ is obtained by contacting
 the eluate from a ⁶⁸Ge/⁶⁸Ga generator with an anion exchanger and eluting ⁶⁸Ga³⁺ from said anion exchanger.
 - 12. Method according to claim 11 wherein the ⁶⁸Ge/⁶⁸Ga generator comprises a column comprising titanium dioxide.
 - 13. Method according to claims 11 to 12 wherein the anion exchanger comprises HCO₃ as counterions.
- 14. Method according to claims 11 to 13 wherein the anion exchanger is a stronganion exchanger.
 - 15. Method according to claims 6 to 14 for the production of ⁶⁸Ga-radiolabelled PET tracers.

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INTERNATIONAL SEARCH REPORT

		PCT/GB20	PCT/GB2004/001550			
A. CLASSI IPC 7	FICATION OF SUBJECT MATTER A61K51/00 //A61K101:30					
According to	o International Patent Classification (IPC) or to both national classific	ation and IPC				
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Minimum do IPC 7	ocumentation searched (classification system followed by classificat $A61K$	ion symbols)				
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Further documents are listed in the continuation of box C. Patent family members are listed in annex.						
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30 July 2004						
Name and malling address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016		Authorized officer Herrera, S				

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